



Difficulties in the treatment of an Infant with Hemophilia B

Dear Editor,

Hemophilia B is a X-linked recessive bleeding disorder which occurs as a result of Factor IX (FIX) deficiency (1, 2). In some patients with hemophilia, antibody (inhibitor) and allergic reaction may develop against FIX protein which is administered for treatment (3). While antibody develops against the factor administered in approximately 15-25% of the patients with severe hemophilia A, this rate is much lower in patients with hemophilia B (3%). Life-threatening allergic reactions may develop in 50% of the patients who have inhibitor (4).

There is no efficient treatment which can be used to prevent allergic reaction developing secondary to antibody response. In this case report, the difficulties in treatment of an infant with severe hemophilia B who developed antibody and allergic reaction against FIX treatment have been discussed.

Our patient was diagnosed with hemophilia B after investigations performed because of severe hemorrhage which developed following circumcision at the age of five months. At presentation, the bleeding tests revealed an activated partial thromboplastin time (aPTT) of 104 seconds and a FIX level of 1%. A concentrate containing FIX protein obtained from 9 doses of plasma was used to stop bleeding. When the patient was 14-month old, he was rehospitalized because of head trauma. His cranial ultrasonography and brain tomography findings were found to be normal. FIX concentrate (Replene®) was initiated at a dose of 80 IU/kg by way of slow intravenous infusion because of head trauma. However, anaphylactic reaction (diffuse urticaria, blushing in the face, cough, dyspnea, cyanosis) developed during infusion of factor

concentrate. Dexamethasone, antihistaminic and adrenaline were administered for treatment. In the following administrations, his treatment was switched to Berinine® which is another FIX concentrate with prophylactic administration of antihistaminic and steroid. A similar anaphylactic reaction developed in the third minute of infusion of this new concentrate and treatment was immediately discontinued. Antihistaminic, dexamethasone and adrenaline were administered again. In the follow-up visit one month later, aPTT was found to be 134 s, the FIX level was found to be 0.1% and the amount of inhibitor was found to be 1.7 BU. As a result of mutation study, CGA>TGA (R252X) mutation was demonstrated in exon 8, n.30875, codon 252. Non-human-derived recombinant active FVIIa (rFVIIa - Novo Seven®) was given for severe bleedings in the following periods because of development of antibody.

Allergic reaction may occur against FIX protein in patients with hemophilia B who have developed antibody in contrast to hemophilia A (5). Follow-up and treatment of hemorrhage is difficult in patients with hemophilia who are allergic against external FIX protein and develop FIX-related antibody. Antibodies are involved in morbidity and mortality related with bleeding. FIX concentrates are not appropriate, because these patients develop antibody and allergy against FIX concentrates. Since treatment with active prothrombin complex concentrate (aPCC) involves FIX, it may result in allergic reaction similarly (6). Development of recombinant FVIIa provided an appropriate treatment option for patients with hemophilia who develop inhibitor. However, no consensus has been made in relation with treatment with aPCC and rFVIIa in FIX patients who have developed antibody (7). In our own individual experience, it was observed that

Address for Correspondence: Serdar Özkasap,

E-mail: sozka1967@yahoo.com

Received: 20.11.2015

Accepted: 01.03.2016

©Copyright 2016 by Turkish Pediatric Association - Available online at www.turkpediatriarsivi.com

DOI: 10.5152/TurkPediatriArs.2016.3626

hemostasis control was provided with aPCC treatment during surgery in patients with hemophilia B who developed antibody (8). It should be kept in mind that the cost of recombinant FVIIa concentrates is high and they carry a risk of thrombosis. Therefore, inhibitor elimination and desensitization treatment should be considered in these patients (9). Elimination of antibody is considerably difficult and treatment success is poor in patients with hemophilia B who are allergic against Factor IX concentrates and who develop antibody. Immune tolerance induction (ITI) treatment which is commonly used in patients with hemophilia A to eliminate antibody and is successful is generally ineffective in these patients. It is recommended that patients with high-risk in terms of development of antibody (patients with complete gene loss) should be identified with molecular analyses at the time of first diagnosis and monitored closely in the beginning of treatment (5). In patients with hemophilia B, it is recommended that the first 10-20 treatments should be administered in hospital in terms of allergy and family members should be trained in terms of the risk of anaphylaxis (10).

In conclusion, we aimed to emphasize the difficulties in follow-up of patients with severe hemophilia B who develop allergic reaction and inhibitor against FIX concentrates. Since life-threatening allergic reactions against Factor IX concentrates may develop, the initial administrations in treatment of patients with newly diagnosed hemophilia B should be definitely performed in hospital environment. Recombinant FVIIa is the only appropriate option for treatment of hemorrhage in these patients. Despite a need for inhibitor elimination and desensitization treatment, there is no current application in this area and further studies are needed.

References

1. Soucie JM, Jackson D, Evatt B. Occurrence of hemophilia in the United States: The Hemophilia Surveillance System Project Invest. Am J Hematol 1998; 59: 288-94. [\[CrossRef\]](#)
2. Iorio A, Olioecchio E, Morfini M, et al. Italian Registry of Haemophilia and Allied Disorders. Objectives, methodology and data analysis. Haemophilia 2008; 14: 444-53. [\[CrossRef\]](#)
3. DiMichele DM. Inhibitor treatment in haemophilias A and B: Inhibitor diagnosis. Haemophilia 2006; 12: 37-41. [\[CrossRef\]](#)
4. DiMichele D. Inhibitor development in haemophilia B: an orphan disease in need of attention. Br J Haematol 2007; 138: 305-15. [\[CrossRef\]](#)
5. Warrier I, Lusher JM. Development of anaphylactic shock in haemophilia B patients with inhibitors. Blood Coagul Fibrinolysis 1998; 9: 125-8.
6. Schoppmann A, Jaeger K, Berg R, et al. Review of the literature of feiba administration in patients with hemophilia B and inhibitors. J Coagul Disord 2011; 3: 14-26.
7. Ewenstein BM, Wong WY, Schoppmann A. Bleeding reduction during secondary prophylaxis with bypassing agents in inhibitor patients. Thromb Res 2011; 127: 174-5. [\[CrossRef\]](#)
8. Zülfikar B, Kavaklı K, Antmen B, Atay A, Şalcıoğlu Z, Ar C. Hemostatic management of surgical procedures with Feiba: a Turkish case series. Haemophilia 2008; 14 (Suppl 2): 54-5.
9. Verma D, Moghimi B, LoDuca PA, Singh HD, Hoffman BE, Herzog RW, Daniell H. Oral delivery of bioencapsulated coagulation factor IX prevents inhibitor formation and fatal anaphylaxis in hemophilia B mice. Proc Natl Acad Sci U S A 2010; 13: 107: 7101-6. [\[CrossRef\]](#)
10. Warrier I, Ewenstein BM, Koerper MA, et al. Factor IX inhibitors and anaphylaxis in hemophilia B. J Pediatr Hematol Oncol 1997; 19: 23-7. [\[CrossRef\]](#)

Serdar Özkasap¹, Selim Dereci², Gül Nihal Özdemir^{3,4}, Bülent Zülfikar^{3,4}

¹Division of Hematology and Oncology, Department of Pediatrics, Recep Tayyip Erdoğan University School of Medicine, Rize, Turkey

²Division of Pediatric Gastroenterology, Department of Pediatrics, Süleyman Demirel University School of Medicine, Isparta, Turkey

³Division of Hematology and Oncology, Department of Pediatrics, İstanbul University Cerrahpaşa School of Medicine, Rize, Turkey

⁴Hemophilia Society of Turkey