See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/228838278

Chemical Constituents and Antimicrobial Activity of the Essential Oil from Vicia dadianorum Extracted by Hydro and Microwave Distillations

Article in Records of Natural Products · January 2012





Rec. Nat. Prod. 6:1 (2012) 49-56

records of natural products

Chemical Constituents and Antimicrobial Activity of the Essential Oil from *Vicia dadianorum* Extracted by Hydro and Microwave Distillations

Nuran Kahriman¹, Büşra Yaylı¹, Murat Yücel¹, Şengül Alpay Karaoğlu² and Nurettin Yaylı^{3*}

¹Department of Chemistry, Faculty of Science, Karadeniz Technical University, 61080, Trabzon,

Türkiye

²Department of Biology, Faculty of Arts and Sciences, Rize University, 53100, Rize, Türkiye ³Faculty of Pharmacy, Karadeniz Technical University, 61080, Trabzon, Türkiye

(Received May 21, 2010; Revised August 25, 2010; Accepted August 31, 2010)

Abstract: The aim of this research was to investigate the influence of extraction methods on yield and chemical composition of the essential oil of *Vicia dadianorum* Somm. & Lev. The volatiles of *V. dadianorum* have been isolated by hydro and microwave distillations (HD and MD). The compositions of the essential oils were characterized by GC-FID and GC-MS. A total of seventy-six and fifty-six compounds were identified, constituting over 90.9%, and 80.1% of oil composition of *V. dadianorum*, respectively. Sesquiterpene hydrocarbons were shown to be the main group of volatiles (HD: 26.2% and MD: 15.9%). The major terpene constituent of the essential oils of *V. dadianorum* was γ -elemene (HD, 13.7% and MD, 8.4%). Comparative study showed that the amount of total volatiles (90.9%) and the major constituent (26.2%) were found to be better in HD of *V. dadianorum*. The antimicrobial activity of the isolated essential oils of the plant was also investigated, and it showed moderate antimicrobial and antifungal activities against *Staphylococcus aureus*, *Enterococcus faecalis*, *Bacillus cereus*, *Mycobacterium smegmatis* and *Candida albicans*.

Keywords: *Vicia dadianorum,* hydrodistillation; microwave distillation; essential oil, antimicrobial activity; GC-MS.

1. Introduction

The genus *Vicia* L. (Leguminosae) is represented with 61 native species including 87 intraspecific taxa, 5 of them is endemics in Turkey [1-3]. *Vicia* is a medium sized genus but it is economically considerable genus because of the two early domesticated plants: *V. faba* and *V. ervilia* [4]. This genus includes some food crops and forage plants such as *V. sativa* cultivated in many countries including Turkey [5]. *V. dadianorum* Somm. & Lev. is a perennial herb with a creeping rootstock and it is grown

^{*} Corresponding author: E-Mail: yayli@ktu.edu.tr; Phone:+90-462-3772486 Fax:+90-462-3253196

in subalpine meadows. It is a Euxine element and mainly distributed in NE Anatolia [1].

Essential oils in plant are complex volatile mixtures exist at low concentrations. Before analyzing the oils, they have to be extracted from the plant. Several extraction processes have been used in order to obtain the high yield of components [6]. The effect of different distillation methods on oil content and composition of aromatic plants have been previously mentioned [7-9]. Recently, a microwave distillation has been developed for extracting volatile products [10-14].

V. faba has been investigated for volatiles [15-16]. However, no published study has previously reported on the essential oil composition of *V. dadianorum* grown in Turkey. The present study was designed to analyze the chemical composition and compare the essential oil contents of *V. dadianorum* extracted by hydrodistillation and a microwave distillation as well as to evaluate their antimicrobial activity.

2. Materials and Methods

2.1. Plant Material

V. dadianorum was collected in Yağmurdere valley, Gümüşhane-Turkey (at heights of ~2020 m) in the northeastern part of Turkey in May, 2010. The plant was authenticated by Prof. S. Terzioğlu [1-4]. Voucher specimen was deposited in the Herbarium of the Faculty of Forestry, KATO (KATO: 12171), Karadeniz Technical University, Turkey.

2.2. Hydrodistillation Apparatus and Procedure

The fresh plant material (150 g) were grounded into small pieces and submitted to hydrodistillation (HD) using a Clevenger-type apparatus with cooling bath (-15 °C) system (4h) (yield (v/w): 0.035%). The obtained oil was extracted with HPLC grade n-hexane (0.5 mL) and dried over anhydrous sodium sulphate and stored at -5 °C in a sealed brown vial.

2.3. Microwave Distillation Apparatus and Procedure

Microwave distillation (MD) was performed at atmospheric pressure with a Milestone DryDIST microwave apparatus using a fixed power of 750 W for 40 min. Temperature was monitored by an external Infrared (IR) sensor. The fresh plant material (150 g) were grounded into small pieces, then placed in a round bottom flask (2 L) with 50 ml water and submitted to microwave distillation (MD) using a Clevenger-type apparatus with cooling bath (-15 °C) system (yield (v/w): 0.055%). The obtained oil was extracted with HPLC grade n-hexane (0.5 mL) and dried over anhydrous sodium sulphate and stored at -5 °C in a sealed brown vial.

2.4. Gas Chromatography (GC) and Gas Chromatography-Mass Spectrometry (GC-MS) Analysis

GC-FID and GC-MS analyses were done as described previously [14].

2.5. Identification of Constituents

The components of the oils were identified by comparison of their mass spectra with those of a computer library or with authentic compounds (α -pinene, α -terpineol, linalool, pentadecane, heptadecane, heneicosane, docosane, tricosane, tetracosane, and pentacosane) and confirmed by comparison of their retention indices, either with those of authentic compounds or with data published in the literature [17-28].

2.6. Antimicrobial Activity Assessment

All test microorganisms were obtained from the Hifzissihha Institute of Refik Saydam (Ankara, Turkey) and were as follows: *Escherichia coli* ATCC 25922, *Yersinia pseudotuberculosis* ATCC 911, *Pseudomonas auroginosa* ATCC 27853, *Staphylococcus aureus* ATCC 25923, *Enterococcus faecalis* ATCC 29212, *Bacillus cereus* 709 ROMA, *Mycobacterium smegmatis* ATCC607 and *Candida albicans* ATCC 60193. All the essential oils were dissolved in hexane to prepare chemicals stock solution of 9.000-21.100 µg /400 µL.

2.7. Agar Well Diffusion Method

Simple susceptibility screening test using agar-well diffusion method [29] as adapted earlier [30] was used. Each bacterium was suspended in Mueller Hinton (MH) (Difco, Detroit, MI) broth. The yeast like fungi was suspended in Yeast extracts broth. Then the microorganisms were diluted approximately 10^6 colony forming unit (cfu) per mL. For yeast like fungi, Sabouraud Dextrose Agar (SDA) (Difco, Detriot, MI) were used. Brain Heart Infusion Agar (BHI) (Difco, Detriot, MI) was used for *M. smegmatis*. They were "flood-inoculated" onto the surface of MH and SD agars and then dried. Five-millimeter diameter wells were cut from the agar using a sterile cork-borer, and 40 μ L of the extract substances were delivered into the wells. The plates were incubated for 18 h at 35°C. The *M. smegmatis* was grown for 3 days on BHI agar plates at 35 °C [31]. Antimicrobial activity was evaluated by measuring the zone of inhibition against the test organism. Ampicillin (10 μ g), Streptomisin (10 μ g) and fluconazole (5 μ g) were standard drugs. Hexane was used as solvent control. The results are shown in table 2.

3. Results and Discussion

The volatile oils obtained after hydrodistillation and microwave distillation of the *V*. *dadianorum* gave an average yields (v/w) of 0.035% and 0.055%, respectively. In total, GC-MS analyzes allowed the identification of 86 volatile compounds [17-28] (76, HD and 56, MD), accounting for 90.0% and 80.1% of the detected GC peak areas, respectively. The list of the identified volatile constituents as well as their grouping into nine classes, namely monoterpene hydrocarbons, oxygenated monoterpens, sesquiterpene hydrocarbons, oxygenated sesquiterpenes, terpene related compounds, aldehydes, esters, hydrocarbons and others with the ratios is given in table 1. The higher number of compounds extracted by HD (76 components) compared to MD (56 components) is probably related to the possible degradation of products by oxidation or hydrolysis, because a longer extraction time (3 h for HD and 30 min for MD) and a greater quantity of water (2 L for HD and 50 ml for MD) used.

Sesquiterpene hydrocarbones were found as the major group of compounds in *V. dadianorum*, constituted 26.2% in HD and 15.9% in MD of the oils. Among them, γ -elemene (13.7% HD and 8.4% MD), (*Z*)-caryophyllene (2.6% HD and 1.5% MD), and β -elemene (2.6% HD and 1.1% MD) were identified as the main components (Figure 1).

Essential oil from Vicia dadianorum



Figure 1. Main sesquiterpene components in the essential oils of V. dadianorum

Linalool (6.5%) was the major compound of the oxygenated monoterpenes (1.6% HD and 1.1%)MD). V. dadianorum oils were characterized by high content of aldehydes (17.4% HD and 19.8% MD), esters (10.0% HD and 19.2% MD), and terpene related compounds (9.8% HD and 8.4% MD). Among the aldehydes, (2E,4E)-decadienal (4.5% HD and 6.0% MD), nonanal (3.1% HD and 2.1% MD), (2E)-nonen-1-al (2.9% HD and 1.9% MD) were found to be the main constituents. Furthermore, hexahydro farnecylacetone (4.7% HD and 5.1% MD) was determined as the major constituent of the terpene related compounds. Monoterpene hydrocarbons were determined only in the oil of HD. The MD oil could be distinguished from the HD oil by their richness in aldehydes (17.4% HD and 19.8% MD) and hydrocarbons (10.0% MD and 19.2% HD). The HD oil could be differentiated from the MD oil by their greater richness in sesquiterpene hydrocarbones (26.2% HD and 15.9% MD), oxygenated monoterpenes (2.7% HD and 1.7% MD), and monoterpene hydrocarbones (1.4% HD and 0% MD). The numbers of the identified terpenoids in the oils of V. dadianorum were 38 (HD) and 16 (MD) compounds. The analysis of variance showed that the distillation method had a significant effect on the oil content of V. dadianorum. The highest oil yield was obtained by microwave distillation method. This may be due to fact that in microwave distillation, type and situation of plant material. mode of charging, and grade of insulation are more important than other distillation method. This result is in agreement with the previous work about the effect of distillation methods on oil contents of volatiles [7-9]. But, comparative study showed that the amount of total volatiles (90.9%) and the major constituent (26.2%) were found to be better in HD of V. dadianorum. Therefore, hydro-distillation could be recommended for the essential oil extraction of V. dadianorum.

Our results appeared to be quite different from previously reported data on the chemical composition of *Vicia* oil since it was devoid of esters, alcohols and oxygenated compounds [15]. The volatile odour chemical of the flowers of the *V. faba* has been identified as (E)- β -ocimene with only trace of other monoterpens [15-16]. Generally, the comparison of our data with the literature showed that the main constituents of chemical composition of the investigated *V. dadianorum* oils were sesquiterpene hydrocarbons and markedly different from known *V. faba* [15-16]. The difference of the composition of the oils could be attributed to the geographical source and the specific climate there.

The antimicrobial activity for the essential oils of *V. dadianorum* was tested *in vitro* using the agar-well diffusion method [29-31] with the microorganisms as seen in Table 2. The essential oils showed moderate antibacterial activity against Gram-positive bacteria *S. aureus*, *E. faecalis*, *B. cereus* 702 Roma, *M. smegmatis*, and pathogenic fungi (*C. albicans*).

Compounds	Area ⁻ (%) HD	Area ⁻ (%)	Ex. RI ^b	Lit. RI
Monoterpene hydrocarbons	110			
α-Pinene ^c	0.4	-	940	939
δ -3-Carene	0.8	_	1031	1031
(E) - β -Ocimene	0.2	-	1051	1050
Oxygenated monoternenes	0.2		1001	1000
Linalool ^c	16	11	1100	1097
<i>a</i> -Terpineol ^c	0.7	0.2	1189	1189
β -Cyclocitral	0.4	0.4	1217	1217
Sesquiternene hydrocarbons	0.1	0.1	1217	1217
<i>a</i> -Copaene	0.2	0.2	1379	1377
(E) - β -Damascenone	0.5	0.3	1382	1385
β -Elemene	2.6	11	1302	1305
β -Longininene	0.2	-	1397	1401
(Z)-Caryonhyllene	2.6	15	1410	1409
(E)-Caryophyllene	0.3	-	1419	1419
β -Duprezianene	-	0.2	1420	1423
v-Flemene	137	8.4	1436	1437
cis-Prenyl limonene	ΩΔ	03	1445	1446
$(F)_B$ -Farnesene	11	0.5	1/5/	1/157
(<i>L</i>)- <i>p</i> -ramesene	1.1	0.7	1454	1457
a-Actionatione	0.1	0.0	1408	1400
β Solinono	0.7	0.9	1400	1400
$(F) \in \mathcal{B}$ Discholong	0.2	0.3	1490	1490
(E) - ρ -Disabolence	0.3	0.3	1502	1500
β-Sesquiphenandrene	1.7	1.0	1520	1525
<i>D</i> -Cadillelle	0.2	-	1522	1525
	0.5	-	1551	1550
	1.1	0.7	1301	1301
Use italiaana anavida	0.2		1514	1515
(E) Negatidat	0.3	-	1514	1515
(E)-Nerollana avida	0.2	0.1	1301	1505
Caryophynene oxide	0.0	0.4	1383	1383
$Epi-\alpha$ -cadinoi	0.2	-	1639	1040
Epi- α -mutroioi	0.2	-	1042	1642
Sellina-3, 11-dien-0- α -01	U.3 1 4	-	104/	1044
Jumper campnor (E, E) Estimated	1.0	-	109/	1700
(<i>E</i> , <i>E</i>)-Farnesol	-	0.2	1/2/	1/25
	0.9	0.4	1/39	1/61
repene related compounds	0.4		1056	1050
α -tonene	0.4	-	1230	1256
Dinydroedulan-I	1.9	0.8	1288	1289
1,1,6-1rimethyl-1,2-	0.4	0.1	1351	1354
ainydronaphthalene	<u> </u>	0.2	1450	1 4 5 5
Geranyl acetone	0.4	0.3	1453	1455
(E) - β -lonone	1.7	1.7	1488	1489
Hexahydro farnesylacetone	4.7	5.1	1846	1847
Farnesyl acetone	0.3	0.4	1916	1919
Aldehydes		0.5		
(2E, 4E)-Heptadienal	-	0.6	1014	1015

Table 1. Identified components in the oils of V. dadianorum extracted by HD and MD.

Benzene acetaldehyde	0.3	1.0	1042	1042
(2E)-Octenal	0.5	0.7	1061	1062
Nonanal	3.1	2.2	1101	1101
(2E, 6Z)-Nonadienal	1.5	1.7	1155	1155
(2E)-Nonen-1-al	2.9	1.9	1162	1162
Decanal	0.9	0.4	1204	1202
(2E, 4E)-Nonadienal	-	0.4	1214	1212
(2E)-Decenal	1.7	1.6	1264	1264
(2E.4Z)-Decadienal	0.8	1.9	1291	1293
Undecanal	0.2	_	1304	1307
(2E.4E)-Decadienal	4.5	6.0	1316	1317
3-Dodecen-1-al	0.2	0.3	1362	1359
Dodecanal	0.5	0.5	1408	1409
Tridecanal	-	0.3	1507	1510
Tetradecanal	03	0.3	1612	1613
Fators	0.5	0.5	1012	1015
(7) 2 Hovernul isoprelarate	0.3	0.1	1225	1220
(Z)-5-nexeligitisovalerate	0.3	0.1	1255	1239
(7) 2 Hovernul ticalate	0.3	-	1242	1244
(Z)-3-Hexenyl tiglate	1.0	0.8	1323	1322
Hexyl tiglate	0.2	-	1330	1333
(E)-2-Hexenyl tiglate	0.4	0.2	1338	1338
Benzyl tiglate	0.3	-	1495	1498
Methyl 8-(2-furyl) octanoate	0.1	0.1	1625	1627
Methyl tetradecanoate	-	0.2	1721	1724
Methyl hexadecanoate	0.6	0.8	1922	1922
Ethyl hexadecanoate	0.3	-	1992	1993
Methyl linoleate	0.6	0.5	2094	2096
Methyl linolenate	2.2	-	2102	2101
Methyl octadecanoate	-	0.1	2123	2125
Ethyl linoleate	0.2	0.3	2145	2146
Ethyl linoleolate	0.4	0.5	2158	2161
Hydrocarbons				
Pentadecane ^c	-	0.4	1499	1500
Heptadecane ^c	0.6	0.1	1700	1700
1-Octadecene	0.2	0.2	1789	1790
Heneicosane ^c	2.5	3.1	2099	2100
Docosane ^c	0.9	1.2	2198	2200
Tricosane ^c	27	83	2302	2300
Tetracosane ^c	11	1.0	2400	2400
Pentacosane ^c	2.0	4.9	2500	2500
Others	2.0	ч.)	2300	2500
1 Octan 3 ol	13	17	070	070
2 Octopopo	1.5	1.7	979	979
2 Octanol	0.0	0.0	902	904
3-Octanol 2 Deuted ferrer	-	1.1	988	991
2-Pentyl furan	9.0	2.8	991	993
I-Octanol	0.4	0.6	1070	1068
(6Z)-Nonen-I-ol	-	0.5	11/4	11/1
Hexadecanoic acid	0.3	2.9	1984	1980
			$NC^{u}(HD)$	$NC^{u}(MD)$
Monoterpene hydrocarbons	1.4	-	3	-
Oxygenated monoterpenes	2.7	1.7	3	3
Sesquiterpene hydrocarbons	26.2	15.9	17	3
Oxygenated sesquiterpenes	4.3	1.1	8	4

Terpene related compounds	9.8	8.4	7	6
Aldehydes	17.4	19.8	13	15
Esters	7.5	3.6	13	10
Hydrocarbons	10.0	19.2	7	8
Others	11.6	10.4	5	7
Total	90.9	80.1	76	56

^aPercentages obtained by FID peak-area normalization. ^bRI calculated from retention times relative to that of n-alkanes (C_6 - C_{32}) on the non-polar HP-5 column. ^cIdentified by authentic samples. ^d NC: Number of compounds

Table 2. Screening result for antimicrobial activity of the essential oils from V. dadianorum.

Samulas	Stock	Microorganisms and inhibition zone (mm)							
Samples	(μg/ 400 μL)	Ec	Yр	Pa	Sa	Ef	Bc	Ms	Ca
V. dadianorum (HD)	9.000	-	-	-	8	8	10	30	nt
V. dadianorum (MD)	21.100	-	-	-	10	-	10	30	20
Ampicillin	10	10	10	18	35	10	15	-	-
Streptomycin	10	-	-	-	-	-	-	35	
Fluconazole	5	-	-	-	-	-	-	-	25

Ec: E. coli, Yp: Y. pseudotuberculosis, Pa: P. aeruginosa, Sa: S. aureus, Ef: E. faecalis, Bc: B. cereus 702 Roma, *Ms: M. smegmatis, Ca: C. albicans,* (-): no activity, nt: not tested.

Acknowledgments

We thank Prof.Dr.Salih Terzioğlu for characterization of plant material. This work was supported by grants from Karadeniz Technical University Research Fund and State Planning Agency (DPT) of Turkey.

References

- [1] P.H. Davis (1970). Flora of Turkey and The East Aegean Islands, Vol.3, University Press: Edinburgh.
- [2] N. Maxted (1989). A New Vicia from South-west Turkey. Notes Roy. Bot. Gard., Edinburgh, 453, 453-456.
- [3] J. Zielinski (1992). Vicia parvula (Fabaceae)-A New Species from NW Turkey, The Karaca Arbor. Magazine 1, 121-123.
- [4] P. Hanelt and D. Mettin (1989). Biosystematics of the Genus Vicia L. (Leguminosae), Annual Review of Ecology and Systematics 20, 199-223.
- [5] N. Zeybek and U. Zeybek (1994). *Farmasotik Botanik*, Ege Üniv. Eczacılık Fak. Yayınları, No.2, Ege Üniv. Basımevi, İzmir.
- [6] F. August, A.L. Lopes and C. A. Zini (2003). Sampling and sample preparation for analysis of aromas and fragances *Trends Anal.Chem.* 22, 160-169.
- [7] G.D. Kiran, V. Babu and K. Kaul (2004). Variation in essential oil composition of rose-scented geranium (*Pelargonium* sp.) distilled by different distillation techniques, *Flavour Frag. J.* **20**, 222-231.
- [8] F. Sefidkon, M. Dabiri and A.R. Bidgoly (1999). The effect of distillation methods and stage of plant growth ob the essential oil content and composition of *Thymus kotschyanus* Boiss. & Hohen., *Flavour Frag. J.* 14, 405-408.
- [9] F. Sefidkon, K. Abbasi and G.B. Khaniki (2006). Influence of drying and extraction methods on yield and chemical composition of the essential oil of *Saturaje hortensis*, *Food Chem.* **99**, 19-23.
- [10] M.E. Lucchesi, F. Chemat and J. Smadja (2004). Solvent free microwave extraction of essential oils from aromatic herbs: comparison with conventional hydrodistillation, J. Chrom. A 1043, 323-327.
- [11] M.E. Lucchesi, J. Smadja, S. Bradshaw, W. Louw, and F. Chemat (2007). Solvent free microwave extraction of *Elletaria cardamonum*: a multivariate study of a new technique for the extraction of essential oil, *J. Food Engin.* **79**, 1079-1086.

- [12] N. Asfaw, P. Licence, A.A. Novitskii and M. Poliakoff (2005). Green Chemistry in Ethiopia: the cleaner extraction of essential oils from *Artemisia afra*: a comparison of clean technology with conventional methodology, *Green Chem.* 7, 352-356.
- [13] M.A. Ferhat, B.Y. Meklati, F. Visinoni, M.A. Vian and F. Chemat (2008) Solvent free microwave extraction of essential oils Gree chemistry in the teaching laboratory, *Chem. Today* 26, 48-50.
- [14] N. Kahriman, G. Tosun, H. Genç and N. Yaylı (2010). A comparative essential oil analysis of *Geranium sylvaticum* extracted by hydrodistillation and microwave distillation, *Turk. J. Chem.* 34, 969-976.
- [15] C.J. Sutton, S.J. Keegans, W.D.J. Kirk and E.D. Morgan (1992). Loral volatiles of Vicia faba, Phytochemistry 31, 3427-3428.
- [16] V.K. Rai, S.C. Gupta and B. Singh (2003). Volatile monoterpens from *Prinsepia utilis* L. leaves inhibit stomatal opening in *Vicia fava* L., *Biol. Plantarum* 46, 121-124.
- [17] R.P. Adams (2004). Identification of essential oil components by Gas Chromatography / Quadrupole Mass Spectroscopy, Allured, Carol Stream, IL, USA.
- [18] H.D. Skaltsa, C. Demetzos, D. Lazari and M. Sokovic (2003). Essential oil analysis and antimicrobial activity of eight Stachys species from Greece, *Phytochemistry* 64, 743-752.
- [19] K. Ertugrul, H. Dural, O. Tugay, G. Flamini, P.L. Cioni and I. Morelli (2003). Essential oils from flowers of *Centaurea kotschyi* var. *kotschyi* and *C. kotschyi* var. decumbens from Turkey, *Flavour Frag. J.* 18, 95-97.
- [20] N. Yaylı, A. Yaşar, N. Yaylı, C. Albay, Y. Aşamaz, K. Çoşkunçelebi and Ş. Karaoğlu (2009). Chemical composition and antimicrobial activity of essential oils from *Centaurea appendicigera* and *Centaurea helenioides, Pharm. Biol.* 47, 7-12.
- [21] N. Yaylı, A. Yaşar, C. Güleç, A. Usta, S. Kolaylı, K. Coşkunçelebi and Ş. Karaoğlu (2005). Composition and antimicrobial activity of essential oils from *Centaurea sessilis* and *Centaurea armena*, *Phytochemistry* 66, 1741-1745.
- [22] D.M. Lazari, H.D. Skaltsa and T. Constantinidis (2000). Volatile constituents of *Centaurea pelia* DC., *C. thessala* Hausskn. subsp. *drakiensis* (Freyn & Sint.) Georg. and *C. zuccariniana* DC. from Greece, *Flavour Frag. J.* 15, 7-11.
- [23] N. Yaylı, A. Yaşar, N. Y. İskender, N. Yaylı, T.B. Cansu, K.Coşkunçelebi and Ş. Karaoğlu (2010). Chemical constituents and antimicrobial activities of the essential oils from *Sedum pallidum* var. *bithynicum* and *S. spurium* grown in Turkey, *Pharm. Biol.* 48, 191-194.
- [24] O. Üçüncü, N. Kahriman, S. Terzioğlu, Ş.A. Karaoğlu and N. Yaylı (2010). Composition and antimicrobial activity of the essential oils from flowers of *Senecio othonnae*, *S.racemosus*, and *S.nemorensis*, *Nat. Prod. Com.* 5, 831-834.
- [25] N. Kahriman, C. G. Albay, N. Doğan, A. Usta, Ş.A. Karaoğlu and N. Yaylı (2010). Volatile constituents and antimicrobial activities from flower and fruit of *Arbutus unedo L.*, Asian J. Chem. 22, 6437-6442.
- [26] N. Yaylı, T.B. Cansu, N. Yılmaz, A. Yaşar, M.M. Çetin and N. Yaylı (2010). Constituents of the essential oil from the flower, leaf, and stem of *Salvia viridis* L. grown in Turkey, *Asian J. Chem.* 22, 3439-3446.
- [27] T.B. Cansu, M. Yücel, K. Sinek, C. Baltacı, Ş. A. Karaoğlu and N. Yaylı (2011). Microwave assisted essential oil analiysis and antimicrobial activity of *Myosotis alpestris* subsp. *alpestris, Asian J. Chem.* 23, 1029-1031.
- [28] O. Üçüncü, N. Yaylı, A. Yaşar, S. Terzioğlu and N. Yaylı (2008). Chemical composition of the essential oils from flower, leaf, and stem of *Senecio trapezuntinus* Boiss. grown in Turkey, Natural Product Communications, *Nat. Prod. Com.* 3, 925-928.
- [29] C. Perez, M. Pauli and P. Bazerque (1990). An antibiotic assay by the agar well diffusion method, *Acta Biolo. Med. Exper.* **15**, 113-115.
- [30] I. Ahmad, Z. Mehmood and F. Mohammed (1998). Screening of some Indian medicinal plants for their antimicrobial properties, *J. Ethnophar.* **62**, 183-193.
- [31] G.L. Woods, B.A. Brown-Elliott, E.P. Desmond, G.S. Hall, L. Heifets, G.E. Pfyffer, J.C. Ridderhof, R.J. Jr. Wallace., N.C. Warren and F.G. Witebsky (2003). Susceptibility testing of mycobacteria, nocardiae, and other aerobic actinomycetes, Approved Standard. NCCLS document M24-A, 23, p18.



© 2011 Reproduction is free for scientific studies