

Draft genome sequence of *Serratia proteamaculans* strain SK20 isolated from diseased rainbow trout (*Oncorhynchus mykiss*) in Türkiye

Salih Kumru,¹ Safak Kalindamar²

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT We report the draft genome sequence of *Serratia proteamaculans* strain SK20 isolated from diseased rainbow trout (*Oncorhynchus mykiss*) from an aquaculture facility in Türkiye. The genome is 5.38 Mb with 55% GC content and contains 5,012 predicted protein-coding genes.

KEYWORDS aquaculture, infectious disease, *Serratia proteamaculans*, trout

Serratia members are rod-shaped, Gram-negative, facultatively anaerobic bacteria widely distributed in diverse environments, including soil, water, plants, animals, and aquatic habitats (1–4). While *Serratia marcescens* is the most studied species due to its clinical importance, *Serratia proteamaculans* is recognized for its ecological versatility and opportunistic pathogenicity. This non-pigmented species has been associated with insect diseases, plant interactions, and food spoilage due to its proteolytic and lipolytic activities (5–9). Additionally, virulence factors such as protealysin and OmpX contribute to host interaction and invasion (10, 11). Despite its ecological importance, genomic data on fish-associated isolates remain limited.

Serratia proteamaculans strain SK20 was isolated in June 2011 from liver tissue obtained during routine diagnostic examination of a diseased rainbow trout (*Oncorhynchus mykiss*) from an aquaculture facility in Rize Province, Türkiye (41°01'17.5"N, 40°22'37.5"E). The liver tissue was aseptically processed by homogenization under sterile conditions, serially diluted (10^{-1} to 10^{-6}) in sterile phosphate-buffered saline, and plated on tryptic soy agar. Following incubation at 20°C for 24–48 h, a single colony was selected and purified by repeated streaking. The bacterium was grown aerobically in tryptic soy broth at 20°C with shaking at 150 rpm for 24–48 h. Cells were harvested by centrifugation, and genomic DNA (gDNA) was extracted using the Quick-DNA Fungal/Bacterial Miniprep Kit (Zymo Research) according to the manufacturer's instructions without modification. gDNA quantity and quality were assessed using a Qubit 4.0 fluorometer (Invitrogen) and an Agilent 5400 Fragment Analyzer (Agilent). The sequencing library was prepared using the Nextera XT DNA Library Preparation Kit (Illumina) and sequenced on the Illumina NovaSeq 6000 platform, generating 4,684,275 paired-end reads (2 × 150 bp) (1.4 Gb). Raw read quality was evaluated using FastQC v.0.12.1 (12), and high-quality reads were *de novo* assembled using the CLC Genomics Workbench v.22.0.1 *de novo* assembler (CLC assembler module) with default parameters (similarity fraction: 0.99, word/bubble size: automatic). Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline v.6.10 (13). Default parameters were used for all tools unless otherwise specified.

The assembly resulted in 20 contigs with an N50 value of 553.2 kb, a total genome size of 5,383,469 bp, and an average coverage of 256×. The draft genome contains 5,012 protein-coding genes, 73 tRNAs, and 1 rRNA operon, with approximately 55% GC

Editor Elinne Becket, California State University San Marcos, San Marcos, California, USA

Address correspondence to Salih Kumru, salih.kumru@erdogan.edu.tr.

The authors declare no conflict of interest.

Received 10 March 2026

Accepted 21 April 2026

Published 6 May 2026

Copyright © 2026 Kumru and Kalindamar. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

content. Average nucleotide identity analysis revealed 99.6% similarity to *S. proteamaculans* CCUG 14510 (GenBank: [WBK100000000.1](https://www.ncbi.nlm.nih.gov/nuclot/WBK100000000.1)) (14, 15). Putative antimicrobial resistance genes were identified using the Resistance Gene Identifier v.6.0.5 with the CARD database v.4.0.1 (16). The SK20 genome contains putative genes encoding resistance to benzalkonium chloride, colistin, pulvomycin, tetracycline, cephalosporin, vancomycin, multidrug antibiotics, and beta-lactam antibiotics, based on strict hits $\geq 95\%$ identity with a reference gene in the database. Furthermore, the genome contains putative genes encoding flagella, type I secretion system, type VI secretion system, and type IVa pili, which may contribute to several important processes, such as virulence, colonization, and bacterial competition.

ACKNOWLEDGMENTS

The authors thank Dr. Sevki Kayis and Dr. Akif Er for providing the bacterial isolate.

AUTHOR AFFILIATIONS

¹Faculty of Fisheries, Recep Tayyip Erdogan University, Rize, Türkiye

²Faculty of Art and Science, Ordu University, Ordu, Türkiye

AUTHOR ORCIDS

Salih Kumru  <http://orcid.org/0000-0002-2309-3110>

Safak Kalindamar  <http://orcid.org/0000-0003-2897-9911>

AUTHOR CONTRIBUTIONS

Salih Kumru, Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review and editing | Safak Kalindamar, Investigation, Methodology, Resources, Validation, Visualization, Writing – review and editing

DATA AVAILABILITY

The final draft genome sequence of *Serratia proteamaculans* strain SK20 has been submitted to the National Center for Biotechnology Information (NCBI) genome database under the GenBank accession number [JBSJDP000000000](https://www.ncbi.nlm.nih.gov/nuclot/JBSJDP000000000), BioProject number [PRJNA1255297](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1255297), BioSample number [SAMN48139144](https://www.ncbi.nlm.nih.gov/biosample/SAMN48139144), and SRA [SRR37525866](https://www.ncbi.nlm.nih.gov/sra/SRR37525866).

ETHICS APPROVAL

No experimental procedures were conducted on live animals; therefore, this study is exempt from institutional animal ethics approval.

REFERENCES

- Grimont F, Grimont PAD. 2006. The genus *Serratia*, p 219–244. In Prokaryotes
- Mahlen SD. 2011. *Serratia* infections: from military experiments to current practice. Clin Microbiol Rev 24:755–791. <https://doi.org/10.1128/CMR.00017-11>
- Bernardet JF, Bowman JP. 2015. Bergey's manual of systematics of Archaea and Bacteria
- Austin B, Austin DA. 2016. Bacterial fish pathogens: disease of farmed and wild fish. online resource (XXXV, 732 pages 48 illustrations in color). Springer International Publishing, Cham.
- Nwaiwu O. 2018. Data on evolutionary relationships of *Aeromonas hydrophila* and *Serratia proteamaculans* that attach to water tanks. Data Brief 16:10–14. <https://doi.org/10.1016/j.dib.2017.10.073>
- Williams DJ, Grimont PAD, Cazares A, Grimont F, Ageron E, Pettigrew KA, Cazares D, Njamkepo E, Weill F-X, Heinz E, Holden MTG, Thomson NR, Coulthurst SJ. 2022. The genus *Serratia* revisited by genomics. Nat Commun 13:5195. <https://doi.org/10.1038/s41467-022-32929-2>
- Ercolini D, Russo F, Nasi A, Ferranti P, Villani F. 2009. Mesophilic and psychrotrophic bacteria from meat and their spoilage potential in vitro and in beef. Appl Environ Microbiol 75:1990–2001. <https://doi.org/10.1128/AEM.02762-08>
- Begrem S, Jérôme M, Leroi F, Delbarre-Ladrat C, Grovel O, Passerini D. 2021. Genomic diversity of *Serratia proteamaculans* and *Serratia liquefaciens* predominant in seafood products and spoilage potential analyses. Int J Food Microbiol 354:109326. <https://doi.org/10.1016/j.ijfoodmicro.2021.109326>
- Fougy L, Coeuret G, Champomier-Vergès M-C, Chaillou S. 2017. Draft genome sequence of *Serratia proteamaculans* MFPA44A14-05, a model organism for the study of meat and seafood spoilage. Genome Announc 5:23. <https://doi.org/10.1128/genomeA.00491-17>
- Tsaplina O, Demidyuk I, Artamonova T, Khodorkovsky M, Khaitlina S. 2020. Cleavage of the outer membrane protein OmpX by proteolysin regulates *Serratia proteamaculans* invasion. FEBS Lett 594:3095–3107. <https://doi.org/10.1002/1873-3468.13897>

11. Tsaplina O. 2024. The balance between proteolysis and its substrate, the outer membrane protein OmpX, regulates *Serratia proteamaculans* invasion. *Int J Mol Sci* 25:11. <https://doi.org/10.3390/ijms25116159>
12. Tully B. 2016. Quality Assessment: FastQC v1. *Protocols.io*. <https://doi.org/10.17504/protocols.io.fa3bign>
13. Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of Standard Operating Procedures (SOPs) for (meta)genomic annotation. *OMICS* 12:137–141. <https://doi.org/10.1089/omi.2008.0017>
14. Yoon S-H, Ha S-M, Lim J, Kwon S, Chun J. 2017. A large-scale evaluation of algorithms to calculate average nucleotide identity. *Antonie Van Leeuwenhoek* 110:1281–1286. <https://doi.org/10.1007/s10482-017-0844-4>
15. Rodriguez-R LM, Konstantinidis KT. 2016. The enveomics collection: a toolbox for specialized analyses of microbial genomes and metagenomes. *PeerJ*. <https://doi.org/10.7287/peerj.preprints.1900v1>
16. Alcock BP, Huynh W, Chalil R, Smith KW, Raphenya AR, Wlodarski MA, Edalatmand A, Petkau A, Syed SA, Tsang KK, et al. 2023. CARD 2023: expanded curation, support for machine learning, and resistome prediction at the comprehensive antibiotic resistance database. *Nucleic Acids Res* 51:D690–D699. <https://doi.org/10.1093/nar/gkac920>